## Anti-Influenza Virus Compound from *Streptomyces* sp. RI18

Motoki Takagi,<sup>\*,†</sup> Keiichiro Motohashi,<sup>†</sup> Aya Nagai,<sup>†</sup> Miho Izumikawa,<sup>†</sup> Masahiro Tanaka,<sup>†</sup> Shinichiro Fuse,<sup>‡</sup> Takayuki Doi,<sup>§</sup> Keiichiro Iwase,<sup>II</sup> Atsushi Kawaguchi,<sup>II,⊥</sup> Kyosuke Nagata,<sup>II</sup> Takashi Takahashi,<sup>‡</sup> and Kazuo Shin-ya<sup>\*,#</sup>

Biomedicinal Information Research Center (BIRC), Japan Biological Informatics Consortium (JBIC), 2-4-7 Aomi, Koto-ku, Tokyo 135-0064, Japan, Department of Applied Chemistry, Tokyo Institute of Technology, 2-12-1-H-101 Ookayama, Meguro-ku, Tokyo 152-8552, Japan, Graduate School of Pharmaceutical Sciences, Tohoku University, 6-3 Aza-Aoba, Aramaki, Aoba, Sendai 980-8578, Japan, Department of Infection Biology, Graduate School of Comprehensive Human Sciences, University of Tsukuba, 1-1-1 Tennodai, Tsukuba 305-8575, Japan, Kitasato Institute for Life Sciences, Kitasato University, 5-9-1 Shirokane, Minato-ku, Tokyo 108-8641, Japan, and Biomedicinal Information Research Center (BIRC), National Institute of Advanced Industrial Science and Technology (AIST), 2-4-7 Aomi, Koto-ku, Tokyo 135-0064, Japan

k-shinya@aist.go.jp; motoki-takagi@aist.go.jp

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ABSTRACT



Screening for anti-influenza virus activity in compounds isolated from *Streptomyces* sp. RI18, which was isolated using the membrane filter method, uncovered a novel compound, JBIR-68 (1), which contains a unique skeleton. Its structure was established by extensive NMR and MS analyses. In addition, 1 was synthesized to confirm the configuration of its sugar moiety. Compound 1 inhibited influenza virus growth in plaque assays.

The influenza virus is a major human and animal pathogen with the potential to cause catastrophic mortality. The virus reproduces rapidly, mutates frequently, and occasionally crosses species barriers. The recent emergence in Asia of the avian influenza virus related to a highly pathogenic form of the human virus has highlighted the strong need for effective new treatments.<sup>1-3</sup> Moreover, the emergence and subsequent swift, worldwide spread of the swine-origin influenza virus A (H1N1) in 2009 once again emphasized the urgency for rapidly developing safe, effective vaccines.<sup>4</sup> Potent anti-influenza agents are urgently required in our ongoing fights against seasonal outbreaks and to address the

<sup>&</sup>lt;sup>†</sup> Japan Biological Informatics Consortium (JBIC).

<sup>&</sup>lt;sup>‡</sup> Tokyo Institute of Technology.

<sup>§</sup> Tohoku University.

<sup>&</sup>quot;University of Tsukuba.

<sup>&</sup>lt;sup>⊥</sup> Kitasato University.

<sup>&</sup>lt;sup>#</sup> Advanced Industrial Science and Technology (AIST).

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risk of future pandemics, given the low probability of developing a universally effective influenza virus vaccine in the near future.<sup>5,6</sup> Adamantane derivatives, amantadine and rimantadine, which target M2 ion channels, were the first antiviral drugs used to treat influenza.<sup>7,8</sup> Recently, orally administered oseltamivir and inhaled zanamivir, which target neuraminidase (NA), were also approved for use in treatments.<sup>7,8</sup> However, alternative therapeutics are needed because the appearance of influenza strains resistant to drugs targeting NA, particularly oseltamivir, is an increasing clinical problem.9-11 Thus, new anti-influenza agents are very necessary.

Members of the class Actinobacteria have been extensively studied for their tendency to produce pharmaceutically useful compounds. However, the discovery rate of novel compounds from these strains has decreased significantly. Therefore, we isolated rare Actinobacteria using the membrane filter (MF) method,<sup>12</sup> which allows for the selection of Actinobacteria without antibiotics, unlike traditional selection methods. In fact, we have already isolated a new Streptomyces sp. using this method<sup>13</sup> and have described new promothiocin derivatives from the isolated Streptomyces.<sup>14</sup> In the course of screening for compounds with anti-influenza virus activity from our Actinobacteria cultures, we identified a novel compound from a culture of Streptomyces sp. RI18 (Figure 1). This compound possesses a unique skeleton and was named JBIR-68 (1).



Figure 1. Structure of JBIR-68 (1).

Streptomyces sp. RI18 was cultured for 5 days at 27 °C, with rotary shaking in 500 mL baffled Erlenmeyer flasks each

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containing 100 mL of a production medium. The mycelial cake was extracted with 80% acetone and concentrated in vacuo. The aqueous concentrate was extracted with EtOAc. The EtOAc layer was dried over Na<sub>2</sub>SO<sub>4</sub>, concentrated in vacuo, and subjected to sequential silica gel flash column chromatography. The pure form of 1 was obtained by reversed-phase HPLC (see Supporting Information).

Compound 1 was isolated as a colorless oil  $([\alpha]_D^{25} - 15.0,$ c 0.10, MeOH) that gave a  $[M + H]^+$  ion at m/z 383.2189 in the HRESIMS. This was consistent with the molecular formula  $C_{19}H_{30}N_2O_6$  (calcd for  $C_{19}H_{31}N_2O_6$ , 383.2182). 1 displayed the following IR spectrum: IR (KBr) v<sub>max</sub> 1700 cm<sup>-1</sup>. The <sup>1</sup>H and <sup>13</sup>C NMR spectral data for **1** are shown in Table 1. The structural information for 1 was determined

Table 1. <sup>1</sup>H and <sup>13</sup>C NMR Spectral Data for JBIR-68 (1)<sup>15</sup>

		1
position	<sup>13</sup> C	$^{1}\mathrm{H}$ (J in Hz)
1	67.4	$4.03^{a}$
2	120.7	5.31, t (7.1)
3	140.5	
4	39.5	2.05, m
5	26.2	2.11, m
6	123.8	5.10, t (6.8)
7	131.4	
8	24.7	1.67, s
9	16.6	1.60, s
10	15.3	1.68, s
2'	154.1	
4'	171.7	
5'	30.8	2.60, t (6.8)
6'	36.2	3.60, dt (13.0, 6.8), 3.45, dt (13.0, 6.8)
1″	88.1	5.86, d (6.4)
$2^{\prime\prime}$	71.1	4.14, t (6.4)
3″	71.4	$4.04^{a}$
4‴	83.0	3.96, ddd (6.4, 6.5, 3.2)
5''	69.9	3.57, t (6.4), 3.54, dd (6.5, 3.2)
<sup>a</sup> Overlap	ping.	

from a series of 2D NMR analyses, including HSQC, fieldgradient <sup>1</sup>H-<sup>1</sup>H COSY, and HMBC spectra. Two singlet methyl protons, H-8 ( $\delta_{\rm H}$  1.67) and H-9 ( $\delta_{\rm H}$  1.60), were  ${}^{1}\text{H}-{}^{13}\text{C}$  long-range coupled to each other ( $\delta_{\rm C}$  24.7 and 16.6, respectively), an olefinic quaternary carbon C-7 ( $\delta_{\rm C}$  131.4), and an olefinic methine carbon C-6 ( $\delta_{\rm C}$  123.8). Two <sup>1</sup>H spin systems, the sequence from H-4 to H-6 through H-5, and a correlation between H-1 and H-2, were observed by  ${}^{1}H^{-1}H$ COSY. In addition to these correlations, <sup>1</sup>H-<sup>13</sup>C long-range couplings from a singlet methyl proton H-10 ( $\delta_{\rm H}$  1.68) to an olefinic methine carbon C-2 ( $\delta_{\rm C}$  120.7), an olefinic quaternary carbon C-3 ( $\delta_{\rm C}$  140.5), and a methylene carbon C-4 ( $\delta_{\rm C}$  39.5) revealed a terpene moiety. The high-field <sup>13</sup>C chemical shift value at the methyl carbon C-10 ( $\delta_{\rm C}$  15.3) determined the configuration at C-2 to be E. The  ${}^{13}C$ chemical value at methylene carbon C-1 ( $\delta_{\rm C}$  67.4) created a geraniol residue, as shown in Figure 2.

The sequence from an aminoacetal proton H-1" ( $\delta_{\rm H}$  5.86,  $\delta_{\rm C}$  88.1) to oxymethylene proton H-5" ( $\delta_{\rm H}$  3.57 and 3.54) through oxymethine protons H-2" ( $\delta_{\rm H}$  4.14), H-3" ( $\delta_{\rm H}$  4.04),

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Figure 2. Key correlations observed in COSY (bold lines) and HMBC (arrows) spectra of JBIR-68 (1).

and H-4" ( $\delta_{\rm H}$  3.96) was confirmed by <sup>1</sup>H-<sup>1</sup>H COSY (Figure 2). A <sup>1</sup>H-<sup>13</sup>C long-range coupling from H-1" to an oxymethylene carbon C-4" revealed a ribosyl residue.

The correlation between methylene protons H-5' ( $\delta_{\rm H}$  2.60) and H-6' ( $\delta_{\rm H}$  3.60 and 3.45) was observed in the <sup>1</sup>H-<sup>1</sup>H COSY spectrum. The <sup>1</sup>H-<sup>13</sup>C long-range couplings from H-5' to a carbamide carbon C-2' ( $\delta_{\rm C}$  154.1), an amide carbon C-4' ( $\delta_{\rm C}$ 171.7), and a methylene carbon C-6' ( $\delta_{\rm C}$  36.2), and from H-6' to C-2' and C-4', were observed. The  ${}^{1}H{}^{-13}C$  long-range coupling from H-6' to C-1" revealed a dihydrouridine moiety. The NMR assignment of the dihydrouridine moiety in 1 was supported by the chemical shifts observed by <sup>13</sup>C NMR in the diacetate FR-900848.16 Finally, the connection between the geraniol and the dihydrouridine moieties was elucidated by the <sup>1</sup>H-<sup>13</sup>C long-range couplings from H-1 to C-5" and from H-5" to C-1 (Figure 2). Thus, the structure of 1 was elucidated, as shown in Figure 1. Although FR-900848, isolated from Streptoverticillium fevens HP-891,<sup>17</sup> is the only reported natural compound that possesses the dihydrouridine moiety, 1 is the first compound in which the ribonucleoside and the geranyl residues are connected by an ether bond.

To confirm the absolute configuration of **1**, we synthesized 5'-*O*-geranyl-5,6-dihydrouridine from 2',3'-di-*O*-(*tert*butyldimethylsilyl)uridine (**2**; Scheme 1; also see Supporting



Information).<sup>18</sup> Hydrogenation of **2** in the presence of 10% Pd/C provided 5,6-dihydrouridine (**3**). After formation of the dianion of **3** using NaH and THF, selective *O*-alkylation with geranyl bromide was performed at -78 °C. The subsequent removal of the TBS groups with TBAF and THF yielded 5'-*O*-geranyl-5,6-dihydrouridine, which was purified by silica gel chromatography and by gel permeation chromatography. All resulting spectral data, including the (-) signal of the

optical rotation of the synthetic 5'-O-geranyl-5,6-dihydrouridine (1), were identical to those of JBIR-68 (1). Thus, we confirmed the stereochemistry of JBIR-68 (1).

The ability of **1** to inhibit influenza virus growth was evaluated using plaque assays.<sup>19</sup> Compound **1** (5–50  $\mu$ M) reduced influenza virus plaque number and area (Figure 3;



Figure 3. JBIR-68 (1) inhibits influenza virus plaque formation.

also see Supporting Information). Thus, **1** clearly inhibited the influenza virus growth. Since **1** is a nucleic acid derivative, it was expected to inhibit viral RNA synthesis. However, the mechanism of the antiviral activity of **1** was not clarified (data not shown). Therefore, the target of **1** may be distinct from that of other drugs targeting the influenza virus. A detailed investigation of the mechanism by which **1** inhibits the influenza virus is currently underway.

In conclusion, we isolated a novel compound, **1**, from *Streptomyces* sp. RI18 selected using the MF method. Compound **1** contains a unique structural skeleton and inhibits the influenza virus. We anticipate that this study will convince chemists that *Actinobacteria* species isolated using the MF method can produce bioactive compounds with unique skeletal structures. In addition, since the molecular target of **1** may be different from that of known anti-influenza drugs, it is expected to be a critical compound for developing anti-influenza drugs.

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**Supporting Information Available:** <sup>1</sup>H and <sup>13</sup>C NMR, COSY, HSQC, HMBC, and HRESIMS spectra of **1**. This material is available free of charge via the Internet at http://pubs.acs.org.

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